

1 **Supplementary Materials**

2 *Measurement of soil properties*

3 Soil properties, including soil pH, cation exchange capacity (CEC), and dissolved
4 organic carbon (DOC), were measured in all soil samples. Soil pH was detected with a
5 glass electrode in the soil with a water-to-soil ratio of 2.5:1 (Thermo Orion-868, MA,
6 USA). CEC was measured using BaCl₂ extraction method (ISO11260 1994). DOC was
7 measured in solutions obtained after extraction with borate buffer (pH = 7) using a soil
8 sample to extractant ratio of 1:10 for 1 h and assayed with a total organic carbon (TOC)
9 autoanalyzer (Analytik Jena, Germany). All soil geochemical data and paddy soil
10 attributes in four climatic zones are available in the repository Figshare
11 (<https://doi.org/10.6084/m9.figshare.11493081.v2>).

12 *The N₂O emission potential and its temperature sensitivity*

13 Microcosms were constructed by inoculating 20 g fresh soil in a 250 mL flask. Each
14 soil group which in the sealed flasks were then acclimated under these five temperatures
15 1 day for pre-incubation. After incubation, 2 mL NH₄NO₃ solution (0.00714 mol/L) was
16 applied uniformly to each sample over the soil surface, followed by in an equivalent
17 addition of 10 mg NH₄⁺-N and 10 mg NO₃⁻-N per kg soil. This resulted in a total amount
18 of 0.2 mg NH₄⁺-N and 0.2 mg NO₃⁻-N content in these microcosms. The final soil
19 moisture contents were adjusted to 60% water holding capacity with deionized water.
20 Then, all the flasks of soil samples were sealed and subsequently incubated at the
21 respective temperature in the dark for an additional 9 days. During incubation, the
22 samples were aerated for 5 min each day to maintain an aerobic condition inside the

23 flasks, and the soil moisture content was maintained by adding deionized water every
24 3 days to compensate for the water loss through evaporation. The emission rates of N₂O
25 were measured 1, 2, 5, and 9 days after ammonium nitrate solution application. For
26 each sampling time, a 20 mL gas sample was collected from each flask (after the flasks
27 were sealed with a butyl rubber stopper for 0 and 4 hour) using a gas-tight syringe and
28 transferred to a 50 mL evacuated gas-tight vial for N₂O analysis via gas
29 chromatography. The concentration of N₂O was determined by using of a 3 m (2 mm
30 inner diameter) stainless steel column packed with Porapak Q (80/100 mesh), and an
31 Agilent 7890 gas chromatograph fitted with an electron captured etector set at 300 °C.
32 The column temperature was maintained at 40 °C and the carrier gas was argon–
33 methane (5 %) at a flow rate of 30 mL min⁻¹.

34 The N₂O emission rate was calculated by the following equation:

$$35 F = \rho \times \Delta C \times V \times \frac{273}{(273 + T) \times W} \quad (Eq. 1)$$

36 Where F represents the flux of N₂O ($\mu\text{g N}/(\text{kg}\cdot\text{day})$); ρ is the density of N₂O-N
37 under standard state (g/L); ΔC (ppbV/day or ppmV/day) represents the change of gas
38 concentration after 4-hour incubation; V (mL) is the gas space volume of conical flasks
39 used in present experiment; T (°C) is the incubation temperature, and W (kg) represents
40 the dry weight of soil. For N₂O emission potential, we regarded the soil samples that
41 incubated at 25 °C (the optimal condition of denitrification) during experiment and
42 calculated the N₂O emission rate. For the temperature sensitivity of N₂O emission, the
43 N₂O emission rates (F) were calculated under 5 temperatures (8, 15, 20, 25, and 35 °C)
44 and then calculated the coefficient of variation (CV).

45 The CV was calculated by the following equation:

46

$$CV = \frac{\sigma}{\mu} \quad (Eq. 2)$$

47 Where σ represents the standard deviation and the μ represents the means of the

48 N₂O emission rates.

49 *Gene amplicon sequencing*

50 Soil microbial communities were analyzed by amplicon sequencing of archaea (16S

51 1106F-1378R) (Dubey et al., 2014), bacteria (16S 515F-806R) (Caporaso et al., 2012),

52 fungi (ITS2) (Ihrmark et al., 2012), algae and micro-fauna (18S C4) (Stoeck et al.,

53 2010), respectively. Both forward and reverse primers were tagged with adapter, pad

54 and linker sequences. The barcode sequence (12 mer) was added to reverse primer for

55 pooling multiple samples in one run of MiSep sequencing. All primers were synthesized

56 by Invitrogen (Carlsbad, CA, USA). PCR amplification was performed in triplicate

57 using a Gene Amp PCR-System 9700 (Applied Biosystems, Foster City, CA, USA) in

58 a total volume of 25 µL containing 2.5 µL 10 × PCR bufferIIand 0.5 unit of AccuPrime

59 Taq DNA Polymerase High Fidelity (Invitrogen, Carlsbad, CA, USA), 0.4 µM of each

60 primer, 10 ng template DNA. To target the bacterial 16S rRNA genes, the first step PCR

61 was carried out by primers without the barcode by cycling conditions of initial

62 denaturation at 94 °C for 1 min, and 10 cycles at 94 °C for 20 sec, 53 °C for 25 sec, and

63 68 °C for 45 sec, with a final extension at 68 °C for 10 min. To target the archaeal 16S

64 rRNA genes, PCR was carried out in the same reaction as that for the 16S rRNA gene

65 fragments, and the samples were subjected to 35 cycles of 95 °C for 45 s, 56 °C for 45 s,

66 and 72 °C for 60 s, followed by a final extension at 72 °C for 7 min. For the ITS2 rRNA

67 gene, initial denaturation was at 98°C for 30 s, 32 cycles at 98°C for 10 s, 56°C for 20 s
68 and 72°C for 30 s, with a final extension at 72°C for 8 min. For the 18S rRNA gene,
69 initialization was at 95°C for 5 min, 30 cycles at 94°C for 30 s, 47°C for 45 s and 72°C
70 for 1 min, with a final extension at 72°C for 5 min. The PCR products from three
71 replicates were combined and purified using an Agencourt AMPure XP kit (Beckman
72 Coulter, Brea, CA, USA) following the manufacturer's instructions and eluted in 50 µL
73 deionized water. We used the purified PCR product (15 µL) as the template for the
74 second step PCR amplification through barcoded primers in three technical replicates
75 under the same cycling conditions as the first step except an extension of 10 cycles.
76 PCR products were examined from the second step by electrophoresis with 1% agarose
77 gel. Amplification products of three technical replicates were then combined and
78 quantified by PicoGreen using a FLUOstar Optima (BMG Labtech, Jena, Germany).
79 PCR products were pooled from different samples together with an equal amount and
80 purified through QIAGEN Gel Extraction Kit (QIAGEN Sciences, Germantown, MD,
81 USA) following the manufacturer's instruction and re-quantified by PicoGreen.
82 According to MiSeqTM Reagent Kit Preparation Guide (Illumina, San Diego, CA,
83 USA), the purified mixture was diluted and denatured to obtain 8 pM sample DNA
84 library, and mixed with an equal volume of 8 pM PhiX (Illumina, San Diego, CA, USA).
85 Finally, 600 µL of mixture library was loaded with read 1, read 2 and index sequencing
86 primers (Caporaso et al., 2012) on a 300-cycle (2 × 150 paired ends) kit, and run on a
87 MiSeq at the Institute for Environmental Genomics of the University of Oklahoma.
88 After assigning the sequence to its sample in terms of the barcode, the sequences

were then trimmed using Btrim with threshold of quality scores higher than 20 over a 5 bp window size and a minimum length of 100 bp (Kong, 2011), and pair-end reads were merged into longer reads by FLASH (Magoč & Salzberg, 2011). Unqualified sequences were removed using the following criteria: (i) Phred quality score Q < 20, (ii) reads that shorter than 200 bp, and (iii) reads containing ambiguous residues.. Chimeric sequences were discarded based on the prediction by UPARSE (Edgar, 2013). OTUs were clustered at the 97% similarity level. Final OTUs were generated based on the clustering results, and taxonomic annotations were assigned to each OTU's representative sequence by RDP's Classifier (for 16S rRNA gene, <http://rdp.cme.msu.edu/>), Unite v6 (for ITS rRNA gene, <http://unite.ut.ee/index.php>) and PR2 (for 18S rRNA gene, <https://pr2-database.org/>) database. Each sample was rarefied at 20,000 sequences for the bacterial 16S rRNA gene, 36,984 sequences for archaeal 16S rRNA gene, 10,000 sequences for ITS rRNA gene, and 20,000 sequences for 18S rRNA gene.

103 Network construction

Network analyses were conducted to explore the co-occurrence patterns of microbiomes in paddy soils across four climatic zones based on OTU relative abundances at each field. The relative abundances were calculated as the number of individuals of the total number of archaeal, bacteria, fungal, algal, and micro-faunal sequences, respectively. Covariations were measured across 11 biological replicates of each network to structure a total of 39 networks. Those OTUs which detected more than 4 out of 11 replicate samples were employed for network construction. The Spearman's

111 correlation between two OTUs was estimated using R (version 3.6.0; <http://www.r-project.org/>). Robust correlations were defined as those with Spearman's correlation
112 coefficients > 0.5 and FDR-corrected *P*-values < 0.01. All significant correlations
113 identified here from a pairwise comparison of OTU abundance formed a correlation
114 network.

116 **References**

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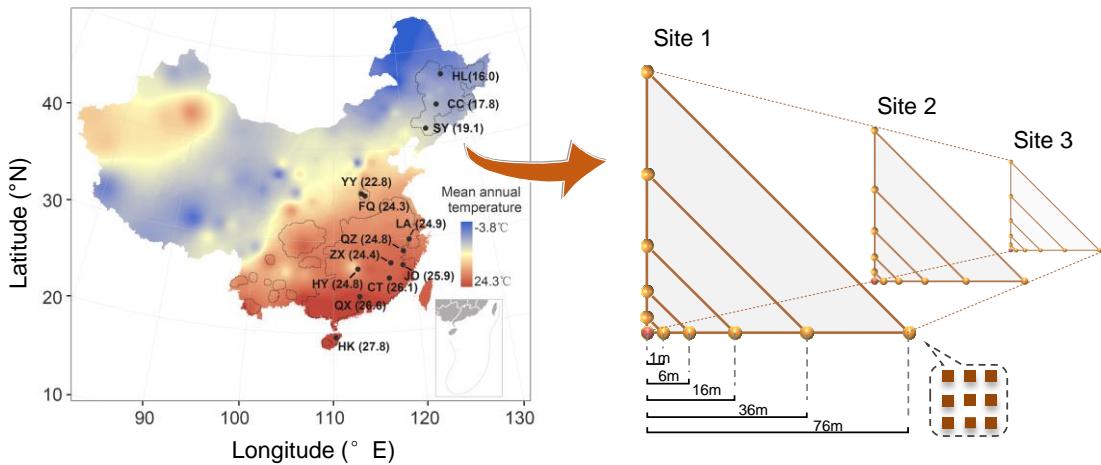


Fig. S1 Sampling locations and strategy. Paddy soil samples were taken from 39 paddy soil sites located in 13 regions of China. Number in parentheses represents mean temperature during the growing season of rice plants ($^{\circ}\text{C}$) in sampling region. Mid-temperate zone includes 3 regions, that is Hailun (HL), Chuangchun (CC), and Shenyang (SY). Warm-temperate zone includes 2 regions, that is Yuanyang (YY) and Fengqiu (FQ). Subtropical zone includes 7 regions, that is Lin'an (LA), Quzhou (QZ), Zixi (ZX), Jian'ou (JO), Changting (CT), Hengyang (HY), and Qingxin (QX). Tropical zone includes 1 region, that is Haikou (HK). At each site, 11 nested samples were collected at distances of 1, 6, 16, 36 and 76 m.

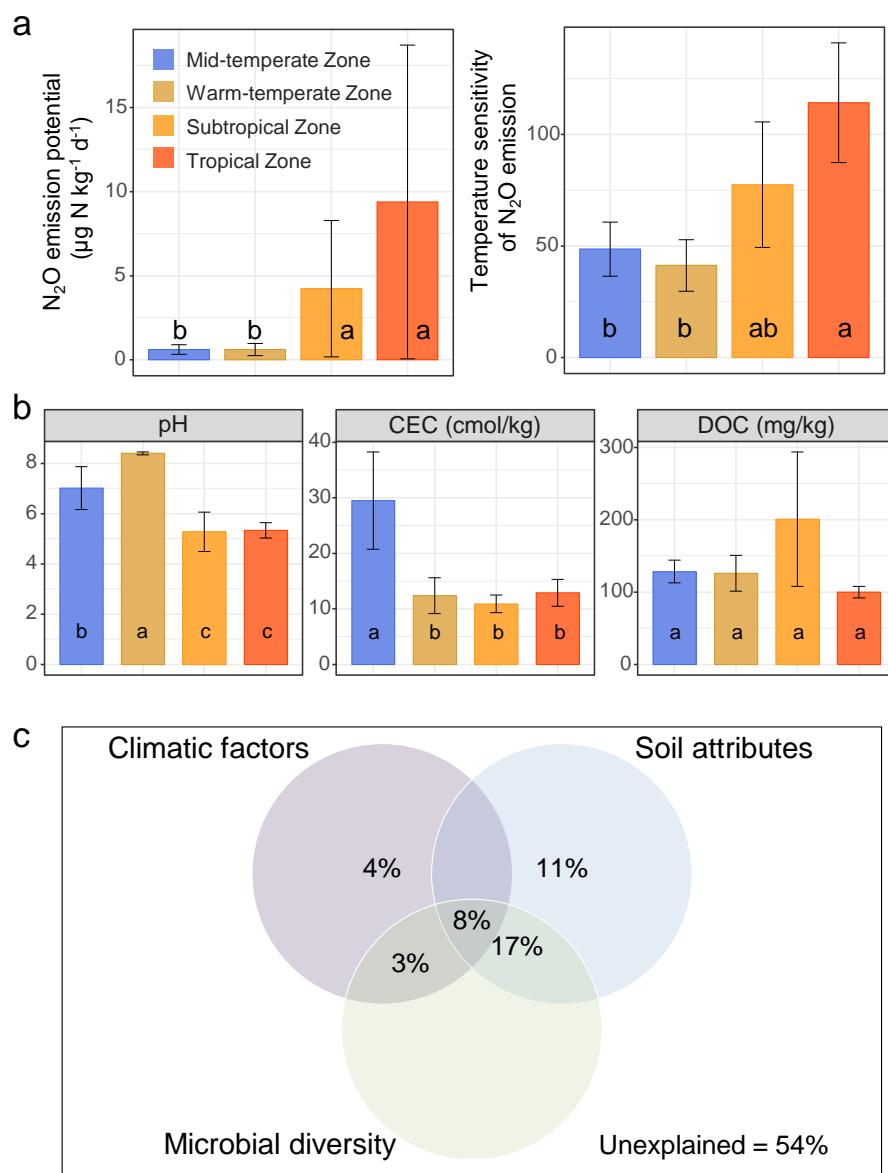


Fig. S2 (a) The N₂O emission potential and the temperature sensitivity, **(b)** soil attributes of the rice paddies in mid-temperate, warm-temperate, subtropical, and tropical zones; **(c)** Variation partitioning analysis of the N₂O emissions and its temperature sensitivity explained by climatic factors, soil attributes, and microbial diversity. Different letters in the panel (a) and (b) indicate significant differences among climatic zones ($p < 0.05$, ANOVA, Tukey HSD). CEC = cation exchange capacity; DOC = dissolved organic carbon. Climatic factors included mean annual temperature and precipitation. Soil attributes included soil pH, CEC, and dissolved organic carbon. The percentage numbers in panel (c) indicated the contributions of different variables to the variation of N₂O emission and its temperature sensitivity quantified by variance partitioning analysis.

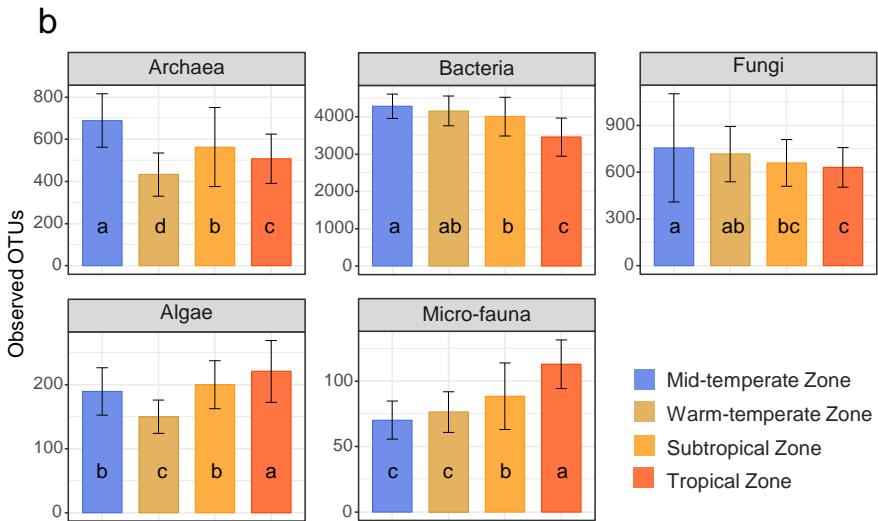
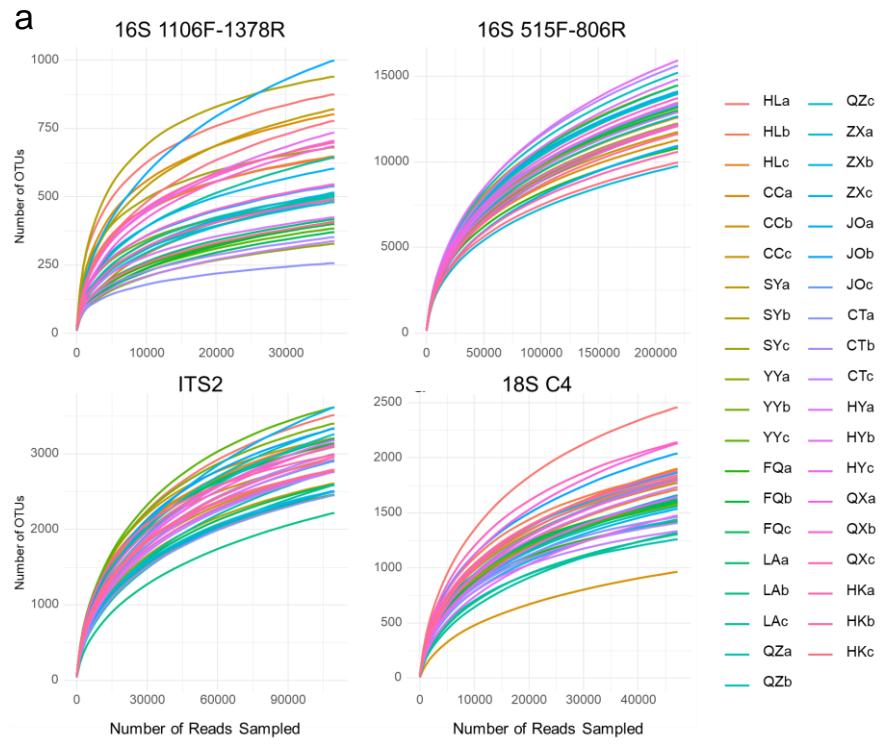


Fig. S3 Rarefaction curves for archaea (16S 1106F-1378R), bacteria (16S 515F-806R), fungi (ITS2), and algae and micro-fauna (18S C4); and the observed OTUs of soil microbiome (archaea, bacteria, fungi, algae, and micro-fauna) of the rice paddies in mid-temperate, warm-temperate, subtropical, and tropical zones.

Different letters in the panel indicate significant differences among different climatic zones ($p < 0.05$, ANOVA, Tukey HSD).

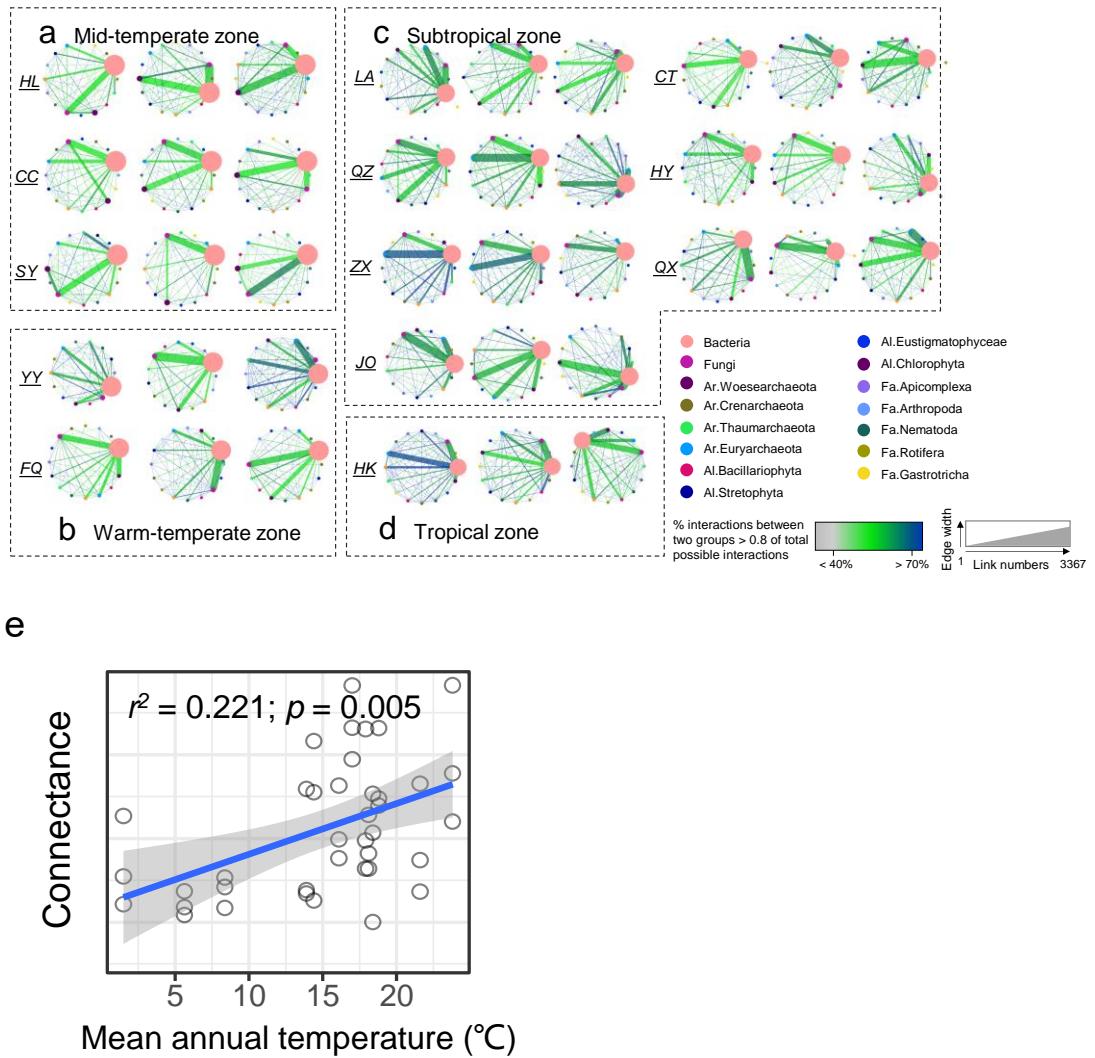
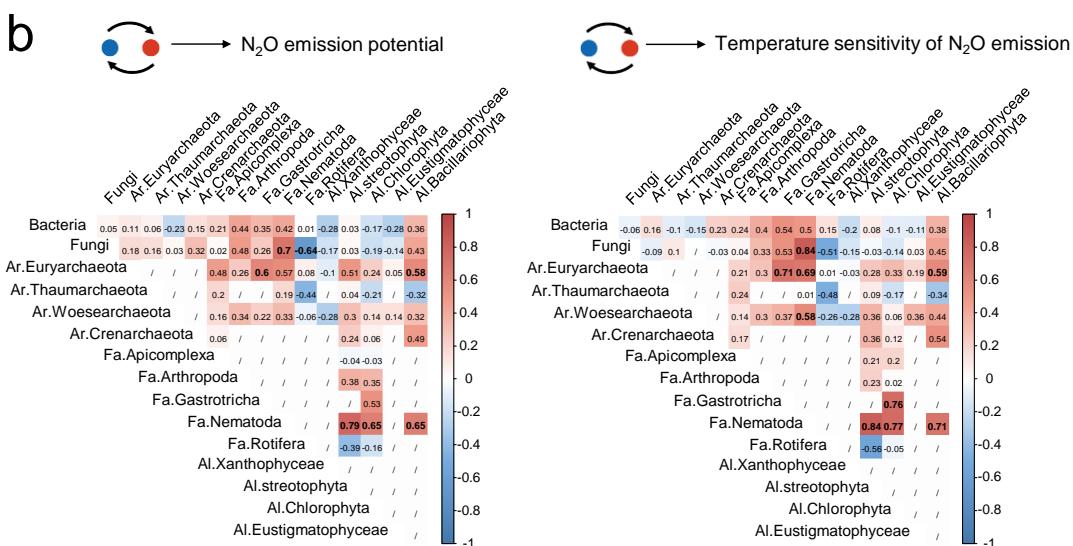
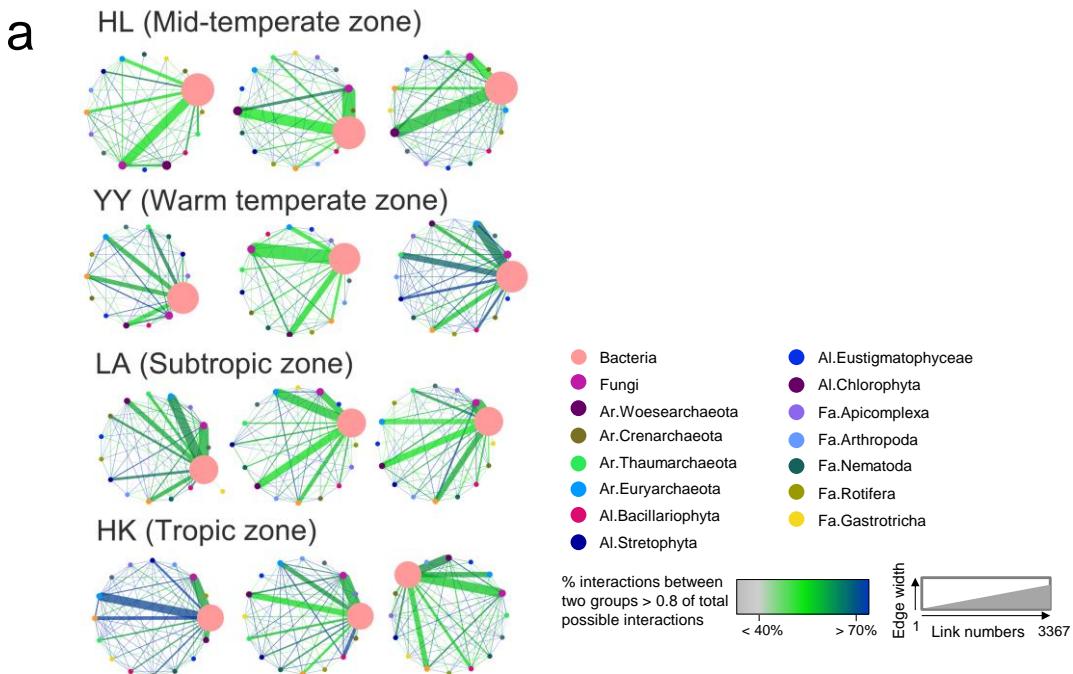


Fig. S4 Network visualization of the interaction strengths between the main microbial groups in 39 paddy soils located in four climatic zones (a-d); The percentage connectance of networks changes with mean annual temperature (e).

Main microbial groups were aggregated by taxonomical classification at the kingdom (bacteria and fungi) or phylum level (archaea and eukaryota). Line width is proportional to the absolute number of correlations > 0.8 , which was divided by the total number of possible interactions to obtain the interaction strength between two groups of species. Line color and transparency are proportional to the interaction strength, as indicated in the legend in the figure. The size of the circles is proportional to the number of species in that group. The connectance of networks in e was calculated by the proportion of correlations > 0.8 divided by the total number of possible interactions. Ar = archaea; Fa = micro-fauna; Al = algae.



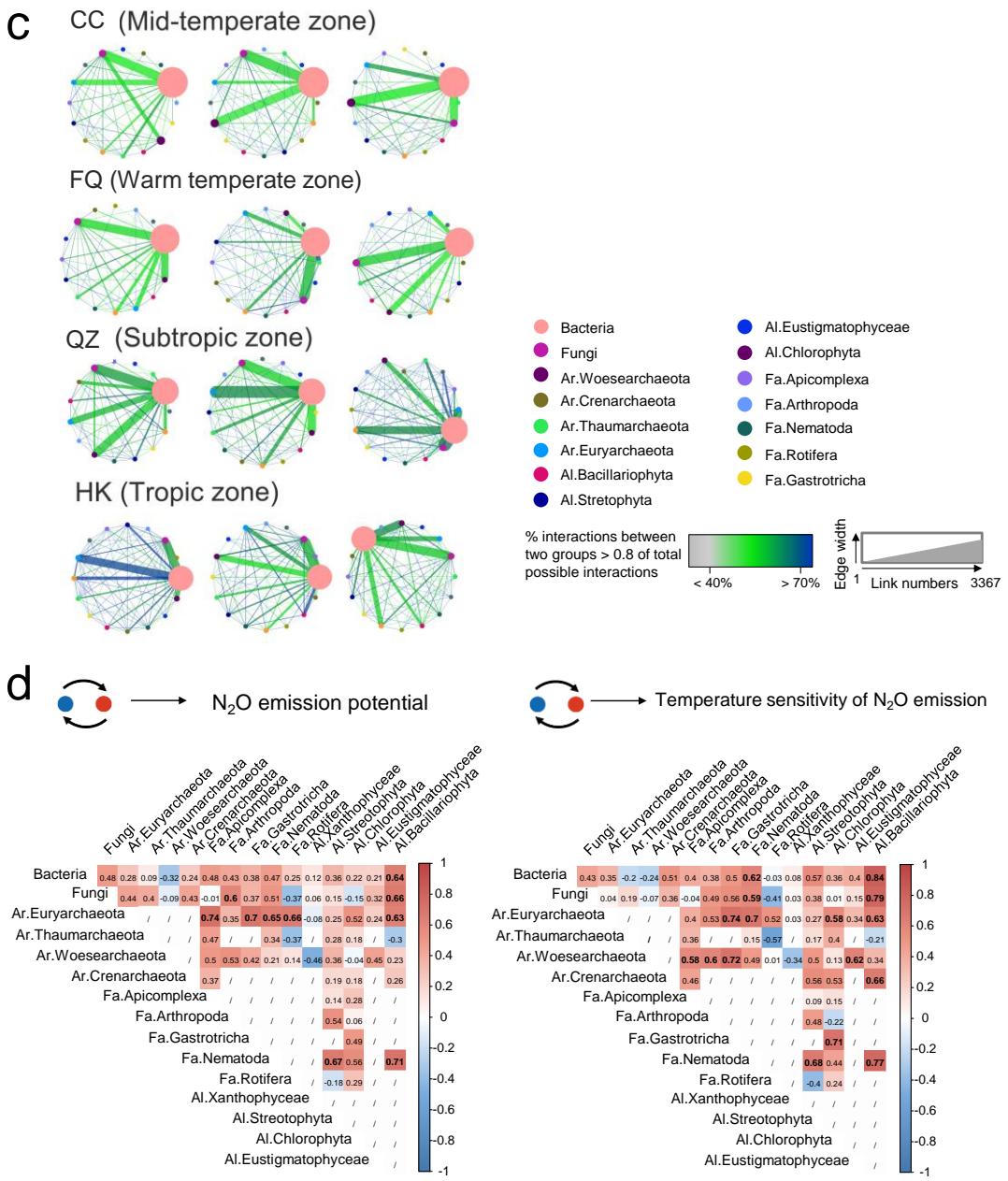


Fig. S5 Relating the main microbial group interaction strengths with N₂O emission in paddy soils using random choosing region from each climatic zone. We first selected the data sets from HL, YY, LA, and HK, respectively from mid-temperate, warm temperate, subtropic, and tropic zones (a, b); we then selected the data sets from CC, FQ, QZ, and HK (c, d). Network visualization of the interaction strengths among the main microbial groups in paddy soils from mid-temperate, warm-temperate, subtropical, and tropical zone (a, c); the correlations between the main group interaction

strengths and the N₂O emission potential and its temperature sensitivity (b, d). Main groups were aggregated by taxonomical classification at the kingdom (bacteria and fungi) or phylum level (archaea and Eukaryotaeukaryota). The proportion of correlations > 0.8 was divided by the total number of possible interactions to obtain the interaction strength between two groups of soil organisms (connectance). Edge width in panel a, c is proportional to the absolute number of correlations > 0.8. Edge color and transparency is proportional to the interaction strength, as indicated in the legend in the figure. The size of the circles is proportional to the number of OTUs in that group. ‘/’ indicate the absence of interactions between these two groups. Significant correlations were indicated marked in bold. Ar = archaea; Fa = micro-fauna; Al = algae.

a Spearman correlations (r) between the dissimilarity of core microbiome and the difference in functional gene abundances determined using Mantel tests.

	N ₂ O emission potential		Temperature sensitivity of N ₂ O emission	
	r	p	r	p
<i>nirK</i>	0.121	< 0.001	0.126	< 0.001
<i>nirS</i>	0.075	< 0.001	0.056	0.003
<i>nosZ</i>	0.146	< 0.001	0.15	< 0.001
<i>nirK/nirS/nosZ</i>	0.129	< 0.001	0.132	< 0.001
Nitrogen cycling genes	0.144	< 0.001	0.157	< 0.001

r and p represent the Mantel test statistic and the p value.

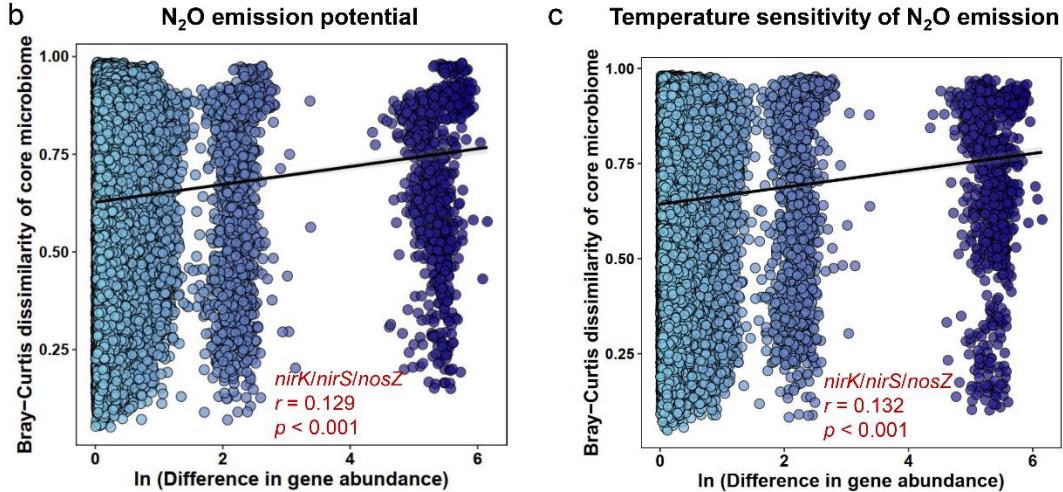


Fig. S6 Linking the core microbiome to nitrogen cycling genes directly involved in N₂O production and reduction. Spearman correlations between the dissimilarity of core microbiome and the log-transformed difference in nitrogen cycling gene abundances determined using Mantel tests (a). Linking the core microbiome related to N₂O emission potential (b) and its temperature sensitivity (c) to nitrogen cycling genes directly contribute to the N₂O pool (i.e., *nirK*, *nirS*, *nosZ*).

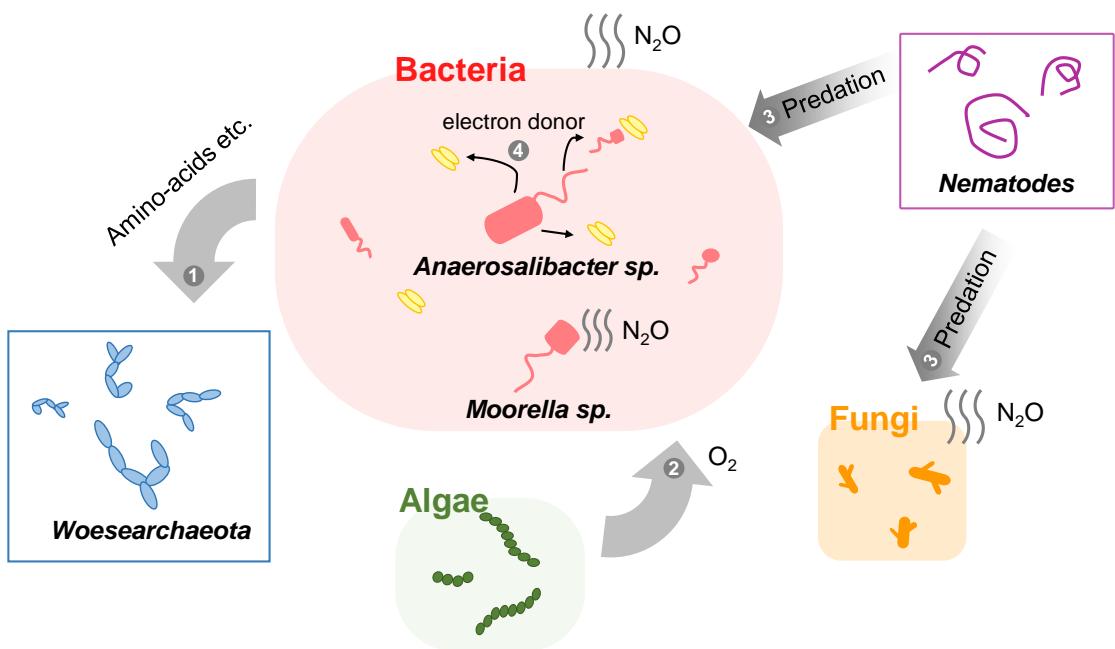


Fig. S7 A pathway diagram visualizing the identified core pair interspecies association in affecting N_2O emission in rice paddy soils according to the two-step method. (1) There might be a potential syntrophic relationship between Woesearchaeota and bacterial members, where bacteria may provide amino-acids and other compounds to compensate for the metabolic deficiencies of Woesearchaeota. (2) Bacteria can break down organic matter using the O_2 produced by photosynthesis of algae, thus providing more electron donor for the denitrification process. (3) Nematode grazing will lead to higher N_2O emissions in soils both indirectly through stimulating microbial activity and directly through excreting N compounds. (4) *Anaerosalibacter* sp. played a central role in enhancing the N_2O emission potential with other bacterial members, which might serve as the provider of electron donor for the denitrification process.

Table S1 Predicting the N₂O emission with environmental factors and microbial interaction strength

Table S1A Multiple linear regressions between environmental factors and the N₂O emission potential or its temperature sensitivity in paddy soils across main rice-cropping areas in China.

	N ₂ O emission potential		Temperature sensitivity of N ₂ O emission	
	Standardized coefficients	p	Standardized coefficients	p
MAT	0.587	0.040	0.785	0.002
pH	-0.249	0.176	-0.470	0.002
CEC	0.335	0.235	0.450	0.048
DOC	0.117	0.470	-0.221	0.090

MAT = mean annual temperature; CEC = cation exchange capacity; DOC = dissolved organic carbon.

Table S1B Summary of the significant regression between MAT and N₂O emission potential and the temperature sensitivity in rice paddies.

	N ₂ O emission potential			Temperature sensitivity of N ₂ O emission		
	r ²	p	AIC	r ²	p	AIC
Linear model	0.19	0.005	227.6	0.34	0.0001	369.6
Exponential model	0.27	0.0006	117.0	0.30	0.0001	42.8
Logarithmic model	0.11	0.04	233.7	0.20	0.005	377.08

The best fit models with lower Akaike information criterion (AIC) and higher r² were marked in bold. Both linear and non-linear regressions were fitted and only significant relationships were shown.

Table S1C The interaction strength (%) between main groups of soil organisms in mid-temperate, warm-temperate, subtropical, and tropical zones in paddy soils across main rice-cropping areas in China.

From	To	Mid-temperate zone	Warm-temperate zone	Subtropical zone	Tropical zone
Bacteria	Ar.Euryarchaeota	52.87	59.92	61.83	67.64
Bacteria	Ar.Thaumarchaeota	60.64	68.04	63.65	80.27
Bacteria	Ar.Woesearchaeota	54.66	55.01	56.72	60.48
Bacteria	Ar.Crenarchaeota	46.97	54.49	61.46	62.56
Bacteria	Fa.Apicomplexa	55.51	58.02	58.90	60.63
Bacteria	Fa.Arthropoda	50.00	59.27	59.48	59.22
Bacteria	Fa.Gastrotricha	55.67	72.82	65.15	66.92
Bacteria	Fa.Nematoda	59.38	58.27	58.28	63.88
Bacteria	Fa.Rotifera	58.71	59.43	52.09	62.81
Bacteria	Al.Xanthophyceae	55.11	49.18	64.31	22.94
Bacteria	Al.streotophyta	60.51	65.10	64.68	72.88
Bacteria	Al.Chlorophyta	50.93	57.01	60.11	62.17
Bacteria	Al.Eustigmatophyceae	52.99	59.55	55.56	57.54
Bacteria	Al.Bacillariophyta	58.20	69.11	61.92	66.15
Bacteria	Fungi	56.29	59.21	60.55	60.26
Fungi	Ar.Euryarchaeota	67.82	68.44	64.73	49.81
Fungi	Ar.Thaumarchaeota	64.90	69.58	70.42	70.62
Fungi	Ar.Woesearchaeota	60.44	60.94	60.77	69.35
Fungi	Ar.Crenarchaeota	56.82	63.11	64.28	37.50
Fungi	Fa.Apicomplexa	66.38	70.71	68.27	63.37
Fungi	Fa.Arthropoda	51.11	70.53	73.51	74.32
Fungi	Fa.Gastrotricha	66.25	85.00	76.02	74.58
Fungi	Fa.Nematoda	76.64	68.53	67.14	72.19
Fungi	Fa.Rotifera	68.24	67.39	62.86	64.29
Fungi	Al.Xanthophyceae	58.16	40.63	66.67	83.33
Fungi	Al.streotophyta	68.04	71.11	71.91	73.91
Fungi	Al.Chlorophyta	59.34	68.62	69.91	62.60
Fungi	Al.Eustigmatophyceae	57.14	63.64	67.67	69.57
Fungi	Al.Bacillariophyta	64.04	74.42	66.40	68.51
Ar.Euryarchaeota	Fa.Apicomplexa	69.86	55.88	66.92	81.40
Ar.Euryarchaeota	Fa.Arthropoda	62.16	45.45	69.39	73.33
Ar.Euryarchaeota	Fa.Gastrotricha	50.00	42.86	75.61	77.50
Ar.Euryarchaeota	Fa.Nematoda	59.46	64.29	62.70	72.07
Ar.Euryarchaeota	Fa.Rotifera	77.14	62.50	60.71	60.00
Ar.Euryarchaeota	Al.Xanthophyceae	60.42	50.00	39.58	86.96
Ar.Euryarchaeota	Al.streotophyta	54.17	70.00	67.23	81.94

From	To	Mid-temperate zone	Warm-temperate zone	Subtropical zone	Tropical zone
Ar.Euryarchaeota	Al.Chlorophyta	60.67	65.22	66.29	93.40
Ar.Euryarchaeota	Al.Eustigmatophyceae	54.55	55.56	57.38	32.89
Ar.Euryarchaeota	Al.Bacillariophyta	62.29	78.70	75.53	77.96
Ar.Thaumarchaeota	Fa.Apicomplexa	85.71	68.75	75.56	64.29
Ar.Thaumarchaeota	Fa.Arthropoda	57.14	77.78	75.76	88.89
Ar.Thaumarchaeota	Fa.Gastrotricha	100.00	/	83.33	60.00
Ar.Thaumarchaeota	Fa.Nematoda	71.43	74.36	72.16	53.33
Ar.Thaumarchaeota	Fa.Rotifera	81.08	77.27	54.55	100.00
Ar.Thaumarchaeota	Al.Xanthophyceae	11.11	0.00	72.22	71.43
Ar.Thaumarchaeota	Al.streotophyta	44.19	80.00	87.55	80.00
Ar.Thaumarchaeota	Al.Chlorophyta	65.44	70.06	101.59	69.89
Ar.Thaumarchaeota	Al.Eustigmatophyceae	0.00	77.78	58.97	/
Ar.Thaumarchaeota	Al.Bacillariophyta	76.92	83.04	74.07	66.67
Ar.Woesearchaeota	Fa.Apicomplexa	65.75	69.35	67.21	64.71
Ar.Woesearchaeota	Fa.Arthropoda	67.86	54.55	70.44	84.44
Ar.Woesearchaeota	Fa.Gastrotricha	64.10	66.67	38.14	48.72
Ar.Woesearchaeota	Fa.Nematoda	60.98	72.32	67.47	68.92
Ar.Woesearchaeota	Fa.Rotifera	71.83	40.00	57.14	28.57
Ar.Woesearchaeota	Al.Xanthophyceae	63.39	50.00	55.32	50.00
Ar.Woesearchaeota	Al.streotophyta	63.33	54.46	67.64	74.40
Ar.Woesearchaeota	Al.Chlorophyta	60.42	59.82	72.98	121.85
Ar.Woesearchaeota	Al.Eustigmatophyceae	58.14	36.36	15.82	60.00
Ar.Woesearchaeota	Al.Bacillariophyta	63.75	68.85	64.86	83.01
Ar.Crenarchaeota	Fa.Apicomplexa	75.00	77.78	80.77	57.14
Ar.Crenarchaeota	Fa.Arthropoda	100.00	50.00	74.19	16.67
Ar.Crenarchaeota	Fa.Gastrotricha	/	0.00	100.00	100.00
Ar.Crenarchaeota	Fa.Nematoda	0.00	33.33	67.57	75.00
Ar.Crenarchaeota	Fa.Rotifera	83.33	100.00	87.50	/
Ar.Crenarchaeota	Al.Xanthophyceae	0.00	/	79.49	100.00
Ar.Crenarchaeota	Al.streotophyta	33.33	50.00	58.22	80.56
Ar.Crenarchaeota	Al.Chlorophyta	43.75	63.64	73.25	71.76
Ar.Crenarchaeota	Al.Eustigmatophyceae	/	40.00	40.74	75.00
Ar.Crenarchaeota	Al.Bacillariophyta	50.00	40.00	76.40	56.25
Fa.Apicomplexa	Al.Xanthophyceae	71.43	100.00	83.33	45.45
Fa.Apicomplexa	Al.streotophyta	83.33	89.29	69.79	62.96
Fa.Apicomplexa	Al.Chlorophyta	74.55	100.00	85.45	83.33
Fa.Apicomplexa	Al.Eustigmatophyceae	100.00	100.00	20.00	100.00
Fa.Apicomplexa	Al.Bacillariophyta	77.78	71.43	47.54	69.57
Fa.Arthropoda	Al.Xanthophyceae	50.00	100.00	100.00	25.00
Fa.Arthropoda	Al.streotophyta	57.14	81.25	60.00	70.00
Fa.Arthropoda	Al.Chlorophyta	56.52	44.44	64.20	33.33

From	To	Mid-temperate zone	Warm-temperate zone	Subtropical zone	Tropical zone
Fa.Arthropoda	Al.Eustigmatophyceae	0.00	50.00	100.00	/
Fa.Arthropoda	Al.Bacillariophyta	40.00	80.00	63.64	100.00
Fa.Gastrotricha	Al.Xanthophyceae	50.00	/	42.86	100.00
Fa.Gastrotricha	Al.streotophyta	33.33	100.00	73.68	50.00
Fa.Gastrotricha	Al.Chlorophyta	54.55	80.00	75.93	72.73
Fa.Gastrotricha	Al.Eustigmatophyceae	/	0.00	62.50	0.00
Fa.Gastrotricha	Al.Bacillariophyta	71.43	/	60.00	51.35
Fa.Nematoda	Al.Xanthophyceae	100.00	100.00	55.56	100.00
Fa.Nematoda	Al.streotophyta	81.82	73.33	82.40	84.62
Fa.Nematoda	Al.Chlorophyta	62.07	70.73	61.36	62.67
Fa.Nematoda	Al.Eustigmatophyceae	60.00	0.00	27.50	40.00
Fa.Nematoda	Al.Bacillariophyta	58.06	80.00	69.23	78.33
Fa.Rotifera	Al.Xanthophyceae	/	/	80.00	0.00
Fa.Rotifera	Al.streotophyta	75.00	100.00	80.00	0.00
Fa.Rotifera	Al.Chlorophyta	65.52	73.33	122.73	75.00
Fa.Rotifera	Al.Eustigmatophyceae	0.00	0.00	100.00	/
Fa.Rotifera	Al.Bacillariophyta	60.00	100.00	62.50	41.67

Note: Darker colors in the cells indicate stronger interaction strength between microbial groups. Ar

= archaea; Fa = micro-fauna; Al = algae.

Table S1D The relationships between within-group interaction strength and the N₂O emission potential and its temperature sensitivity based on Spearman's correlation in rice paddy soils.

		N ₂ O emission potential	Temperature sensitivity of N ₂ O emission
Archaea	R	-0.01	-0.06
	p	0.93	0.72
Bacteria	R	0.01	0.10
	p	0.94	0.54
Fungi	R	0.04	0.20
	p	0.79	0.23
Algae	R	0.23	0.40
	p	0.06	0.02
Micro-fauna	R	-0.24	-0.29
	p	0.14	0.05

Table S2 Pairs of core microorganisms and their independent and cooperative effects on maximizing the N₂O emission in paddy soils based on the two-step criterion.

Table S2A Pairs of core microorganisms and their independent effects on maximizing the N₂O emission potential in paddy soils based on the two-step criterion.

Interaction type	Node A	Node B	B' (A) ^a	B' (B) ^b	C _{ij} ^c	R _{ij} ^d
Between-group	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Halobacteriovorax</i> sp. (Ba.OTU37210)	0.924	0.916	6,441	5,451.299
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Clostridium</i> sp. (Ba.OTU39051)	0.924	0.916	5,700	4,824.277
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Chondromyces</i> sp. (Ba.OTU6099)	0.924	0.916	5,600	4,738.760
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Oxobacter</i> sp. (Ba.OTU5558)	0.924	0.916	5,424	4,591.830
	<i>Halobacteriovorax</i> sp. (Ba.OTU37210)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.916	0.917	4,536	3,807.707
	<i>Halobacteriovorax</i> sp. (Ba.OTU37210)	<i>Methermicoccus</i> sp. (Ar.OTU189)	0.916	0.911	4,368	3,644.259
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Sporotomaculum</i> sp. (Ba.OTU23447)	0.924	0.916	4,180	3,537.063
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Luteitalea</i> sp. (Ba.OTU6324)	0.924	0.915	4,181	3,536.236
	<i>Chondromyces</i> sp. (Ba.OTU6099)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.916	0.917	4,182	3,509.979
	<i>Halobacteriovorax</i> sp. (Ba.OTU37210)	Unclassified <i>Woesearchaeota</i> (Ar.OTU23)	0.916	0.915	4,088	3,426.253
Within-group (Archaea)	<i>Clostridium</i> sp. (Ba.OTU39051)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.916	0.917	4,018	3,372.959
	<i>Halobacteriovorax</i> sp. (Ba.OTU37210)	Unclassified <i>archaeon</i> (Ar.OTU82)	0.916	0.907	3,990	3,315.794
	<i>Oxobacter</i> sp. (Ba.OTU5558)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.916	0.917	3,936	3,304.951
	<i>Clostridium</i> sp. (Ba.OTU39051)	<i>Methermicoccus</i> sp. (Ar.OTU189)	0.916	0.911	3,871	3,229.687
	<i>Oxobacter</i> sp. (Ba.OTU5558)	<i>Methermicoccus</i> sp. (Ar.OTU189)	0.916	0.911	3,792	3,164.567
	<i>Methanocella</i> sp. (Ar.OTU17)	<i>Methanocella</i> sp. (Ar.OTU340)	1	0.649	54	35.042
	<i>Methanocella</i> sp. (Ar.OTU17)	<i>Methanocella</i> sp. (Ar.OTU242)	1	0.444	63	27.978
	<i>Nitrososphaera</i> sp. (Ar.OTU394)	<i>Methanocella</i> sp. (Ar.OTU17)	0.161	1	84	13.519

	<i>Methanocella</i> sp. (Ar.OTU340)	<i>Methanomethylovorans</i> sp. (Ar.OTU321)	0.649	0.410	40	10.631
	<i>Methanocella</i> sp. (Ar.OTU109)	<i>Methanocella</i> sp. (Ar.OTU17)	0.084	1	108	9.078
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Methanobacterium</i> sp. (Ar.OTU160)	0.171	0.072	713	8.764
	<i>Methanocella</i> sp. (Ar.OTU17)	<i>Methanocella</i> sp. (Ar.OTU322)	1	0.113	72	8.135
	<i>Methanosarcina</i> sp. (Ar.OTU50)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.145	0.106	504	7.769
	<i>Methanocella</i> sp. (Ar.OTU17)	<i>Methanocella</i> sp. (Ar.OTU593)	1	0.238	32	7.619
	<i>Methanocella</i> sp. (Ar.OTU242)	<i>Methanocella</i> sp. (Ar.OTU367)	0.444	0.305	54	7.321
Within-group (Bacteria)	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Moorella</i> sp. (Ba.OTU52410)	0.475	0.420	8,000	1,598.890
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	Unclassified bacterium (Ba.OTU52965)	0.475	0.270	8,256	1,059.414
	<i>Moorella</i> sp. (Ba.OTU52410)	<i>Ammoniphilus</i> sp. (Ba.OTU41498)	0.420	0.217	6,903	629.026
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Pyrinomonas</i> sp. (Ba.OTU53333)	0.475	0.265	4,148	521.597
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Sterolibacterium</i> sp. (Ba.OTU11112)	0.475	0.128	7,872	479.746
	Unclassified bacterium (Ba.OTU36668)	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	0.114	0.475	7,298	394.367
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Cnexibacter</i> sp. (Ba.OTU4903)	0.475	0.146	4,664	322.705
	Unclassified bacterium (Ba.OTU52965)	<i>Ammoniphilus</i> sp. (Ba.OTU41498)	0.270	0.217	5,429	317.627
	<i>Haliangium</i> sp. (Ba.OTU24054)	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	0.156	0.475	4,176	309.247
	<i>Haliangium</i> sp. (Ba.OTU24054)	<i>Moorella</i> sp. (Ba.OTU52410)	0.156	0.420	4,386	287.292
Within-group (Fungi)	Unclassified fungus (Fu.OTU1)	<i>Cyathus</i> sp. (Fu.OTU10084)	1	0.390	8	3.122
	Unclassified fungus (Fu.OTU1)	<i>Lobulomyces</i> sp. (Fu.OTU21789)	1	0.050	8	0.402
	Unclassified fungus (Fu.OTU1)	<i>Dominikia</i> sp. (Fu.OTU21962)	1	0.050	8	0.402
	<i>Cyathus</i> sp. (Fu.OTU10084)	Unclassified <i>Zygorhynchus</i> (Fu.OTU27233)	0.390	0.092	4	0.144
	Unclassified <i>Rhizophydium</i> (Fu.OTU4002)	Unclassified <i>Zygorhynchus</i> (Fu.OTU27233)	0.082	0.092	4	0.030
	Unclassified <i>Rhizophydium</i> (Fu.OTU4002)	Unclassified fungus (Fu.OTU10890)	0.082	0.054	4	0.018
	<i>Lobulomyces</i> sp. (Fu.OTU21789)	Unclassified <i>Tremellales</i> (Fu.OTU21826)	0.050	0.017	4	0.003
	<i>Dominikia</i> sp. (Fu.OTU21962)	Unclassified <i>Tremellales</i> (Fu.OTU21826)	0.050	0.017	4	0.003
	Unclassified fungus (Fu.OTU1)	Unclassified fungus (Fu.OTU10224)	1	0	4	0

	<i>Goffeauzyma</i> sp. (Fu.OTU780)	Unclassified <i>Mucorales</i> (Fu.OTU22000)	0.025	0	2	0
Within-group (Algae)	<i>Tetracystis</i> sp. (Al.OTU8)	<i>Chloroparvula</i> sp. (Al.OTU52)	1	0.947	25	23.674
	<i>Tetracystis</i> sp. (Al.OTU8)	<i>Pseudopleurococcus</i> sp. (Al.OTU89)	1	0.318	15	4.773
	<i>Nannochloris</i> sp. (Al.OTU43)	<i>Chloroparvula</i> sp. (Al.OTU52)	0.365	0.947	8	2.766
	<i>Nautococcus</i> sp. (Al.OTU32)	<i>Pterosperma</i> sp. (Al.OTU388)	0.536	0.279	15	2.245
	<i>Chloroparvula</i> sp. (Al.OTU52)	<i>Pterosperma</i> sp. (Al.OTU388)	0.947	0.279	8	2.114
	<i>Tetracystis</i> sp. (Al.OTU8)	<i>Crustomastix</i> sp. (Al.OTU100)	1	0.156	6	0.937
	<i>Nephroselmis</i> sp. (Al.OTU34)	<i>Nautococcus</i> sp. (Al.OTU32)	0.101	0.536	12	0.650
	<i>Monomastix</i> sp. (Al.OTU180)	<i>Nautococcus</i> sp. (Al.OTU32)	0.101	0.536	12	0.650
	<i>Tetracystis</i> sp. (Al.OTU8)	<i>Nephroselmis</i> sp. (Al.OTU34)	1	0.101	6	0.606
	<i>Tetracystis</i> sp. (Al.OTU8)	<i>Monomastix</i> sp. (Al.OTU180)	1	0.101	6	0.606
Within-group (Micro-fauna)	Unclassified eukaryote (Fa.OTU12)	<i>Mayamaea</i> sp. (Fa.OTU13)	1	1	1	1
	Unclassified eukaryote (Fa.OTU12)	<i>Phascolodon</i> sp. (Fa.OTU23)	1	1	1	1
	<i>Mayamaea</i> sp. (Fa.OTU13)	<i>Phascolodon</i> sp. (Fa.OTU23)	1	1	1	1
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	Unclassified eukaryote (Fa.OTU12)	0	1	2	0
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Mayamaea</i> sp. (Fa.OTU13)	0	1	2	0
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Enchelyodon</i> sp. (Fa.OTU16)	0	1	4	0
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Phascolodon</i> sp. (Fa.OTU23)	0	1	2	0
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Prodorylaimus</i> sp. (Fa.OTU26)	0	0	1	0
	Unclassified eukaryote (Fa.OTU12)	<i>Prodorylaimus</i> sp. (Fa.OTU26)	1	0	2	0
	<i>Mayamaea</i> sp. (Fa.OTU13)	<i>Prodorylaimus</i> sp. (Fa.OTU26)	1	0	2	0

^{a, b} denotes the species keystone ness after standardizing (Eq. 4)

^c represents the number of checkboard units based on Eq. 7

^d represents their independent effects on maximizing the function calculated by Eq. 11

Table S2B Pairs of core microorganisms and their cooperative effects on maximizing the N₂O emission potential in paddy soils based on the two-step criterion.

Interaction type	Node A	Node B	B' (A) ^a	B' (B) ^b	T _{ij} ^c	R _{ij} ^d
Between-group	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Methanobacterium</i> sp. (Ar.OTU13)	0.950	0.930	54,696	48,337.783
	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Methanocella</i> sp. (Ar.OTU32)	0.950	0.942	53,193	47,598.570
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Nautococcus</i> sp. (Al.OTU32)	0.924	0.916	50,550	42,779.783
	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Nitrososphaera</i> sp. (Ar.OTU9)	0.950	0.928	46,530	41,019.957
	<i>Nautococcus</i> sp. (Al.OTU32)	Unclassified archaeon (Ar.OTU135)	0.916	0.912	47,790	39,914.262
	<i>Nautococcus</i> sp. (Al.OTU32)	<i>Methanocella</i> sp. (Ar.OTU143)	0.916	0.927	44,058	37,400.229
	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Nitrososphaera</i> sp. (Ar.OTU4)	0.950	0.924	42,517	37,318.420
	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Methanocella</i> sp. (Ar.OTU22)	0.950	0.942	40,677	36,390.424
	<i>Nautococcus</i> sp. (Al.OTU32)	<i>Methanospaerula</i> sp. (Ar.OTU36)	0.916	0.920	37,908	31,932.379
	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Methanocella</i> sp. (Ar.OTU18)	0.950	0.929	33,216	29,325.348
	<i>Nautococcus</i> sp. (Al.OTU32)	<i>Methanococcoides</i> sp. (Ar.OTU74)	0.916	0.920	29,708	25,030.224
	<i>Nautococcus</i> sp. (Al.OTU32)	Unclassified archaeon (Ar.OTU531)	0.916	0.916	25,656	21,515.044
Within-group (Archaea)	<i>Frontonia</i> sp. (Fu.OTU23455)	Unclassified archaeon (Ar.OTU23)	0.917	0.915	25,416	21,318.021
	<i>Frontonia</i> sp. (Fu.OTU23455)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.917	0.917	25,224	21,190.226
	<i>Nautococcus</i> sp. (Al.OTU32)	Unclassified archaeon (Ar.OTU57)	0.916	0.917	24,380	20,466.593
	<i>Methanocella</i> sp. (Ar.OTU17)	<i>Methanocella</i> sp. (Ar.OTU340)	1	0.649	932	604.802
	<i>Methanocella</i> sp. (Ar.OTU17)	<i>Methanocella</i> sp. (Ar.OTU242)	1	0.444	928	412.115
	<i>Methanocella</i> sp. (Ar.OTU17)	<i>Methanocella</i> sp. (Ar.OTU593)	1	0.238	1,175	279.763
	<i>Methanocella</i> sp. (Ar.OTU17)	<i>Methanocella</i> sp. (Ar.OTU162)	1	0.177	1,175	207.645
	<i>Methanocella</i> sp. (Ar.OTU242)	<i>Methanocella</i> sp. (Ar.OTU593)	0.444	0.238	1,428	150.991
	<i>Methanocella</i> sp. (Ar.OTU340)	<i>Nitrosopumilus</i> sp. (Ar.OTU321)	0.649	0.410	474	125.976
	<i>Methanocella</i> sp. (Ar.OTU162)	<i>Methanocella</i> sp. (Ar.OTU242)	0.177	0.444	1,428	112.068

	<i>Methanocella</i> sp. (Ar.OTU340)	<i>Methanocella</i> sp. (Ar.OTU593)	0.649	0.238	708	109.391
	<i>Methermicoccus</i> sp. (Ar.OTU189)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.109	0.106	9,065	104.528
	<i>Methanosphaerula</i> sp. (Ar.OTU10)	<i>Methermicoccus</i> sp. (Ar.OTU189)	0.171	0.109	5,292	98.415
Within-group (Bacteria)	<i>Moorella</i> sp. (Ba.OTU52410)	Unclassified bacterium (Ba.OTU52965)	0.420	0.270	79,106	8,978.741
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Moorella</i> sp. (Ba.OTU52410)	0.475	0.420	39,936	7,981.657
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Pyrinomonas</i> sp. (Ba.OTU53333)	0.475	0.265	54,213	6,817.098
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Ammoniphilus</i> sp. (Ba.OTU41498)	0.475	0.217	61,544	6,340.246
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Denitratisoma</i> sp. (Ba.OTU51233)	0.475	0.157	70,528	5,259.578
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Peredibacter</i> sp. (Ba.OTU54965)	0.475	0.127	81,770	4,955.475
	<i>Moorella</i> sp. (Ba.OTU52410)	<i>Anaerosalibacter</i> sp. (Ba.OTU11112)	0.420	0.128	85,162	4,590.732
	<i>Moorella</i> sp. (Ba.OTU52410)	<i>Gemmata</i> sp. (Ba.OTU8890)	0.420	0.123	83,006	4,307.300
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Cephalothrix</i> sp. (Ba.OTU50224)	0.475	0.131	56,967	3,549.047
	<i>Pyrinomonas</i> sp. (Ba.OTU53333)	<i>Ammoniphilus</i> sp. (Ba.OTU41498)	0.265	0.217	58,459	3,351.579
Within-group (Fungi)	<i>Coemansia</i> sp. (Fu.OTU7779)	Unclassified fungus (Fu.OTU13335)	0	0	28	0
	<i>Coemansia</i> sp. (Fu.OTU7779)	Unclassified Cryptomycota (Fu.OTU21900)	0	0	28	0
	Unclassified fungus (Fu.OTU13335)	Unclassified Cryptomycota (Fu.OTU21900)	0	0	28	0
	<i>Inocybe</i> sp. (Fu.OTU23234)	<i>Acaulospora</i> sp. (Fu.OTU24067)	0	0	28	0
	<i>Rozella</i> sp. (Fu.OTU13113)	<i>Inocybe</i> sp. (Fu.OTU23234)	0.092	0	26	0
	<i>Rozella</i> sp. (Fu.OTU13113)	<i>Acaulospora</i> sp. (Fu.OTU24067)	0.092	0	26	0
	Unclassified fungus (Fu.OTU1)	<i>Cyathus</i> sp. (Fu.OTU10084)	1	0.390	0	0
	Unclassified fungus (Fu.OTU1)	<i>Lobulomyces</i> sp. (Fu.OTU21789)	1	0.050	0	0
	Unclassified fungus (Fu.OTU1)	<i>Dominikia</i> sp. (Fu.OTU21962)	1	0.050	0	0
	<i>Cyathus</i> sp. (Fu.OTU10084)	Unclassified <i>Zygorhynchus</i> (Fu.OTU27233)	0.390	0.092	0	0
Within-group (Algae)	<i>Nannochloris</i> sp. (Al.OTU43)	<i>Chloroparvula</i> sp. (Al.OTU52)	0.365	0.947	43	14.865
	<i>Tetracystis</i> sp. (Al.OTU8)	<i>Crustomastix</i> sp. (Al.OTU100)	1.000	0.156	86	13.431
	<i>Chloroparvula</i> sp. (Al.OTU52)	<i>Pterosperma</i> sp. (Al.OTU388)	0.947	0.279	43	11.363

	<i>Tetracystis</i> sp. (Al.OTU8)	<i>Nephroselmis</i> sp. (Al.OTU34)	1	0.101	86	8.682
	<i>Tetracystis</i> sp. (Al.OTU8)	<i>Monomastix</i> sp. (Al.OTU180)	1	0.101	86	8.682
	<i>Nannochloris</i> sp. (Al.OTU43)	<i>Pterosperma</i> sp. (Al.OTU388)	0.365	0.279	45	4.584
	<i>Nephroselmis</i> sp. (Al.OTU34)	<i>Nautococcus</i> sp. (Al.OTU32)	0.101	0.536	42	2.274
	<i>Monomastix</i> sp. (Al.OTU180)	<i>Nautococcus</i> sp. (Al.OTU32)	0.101	0.536	42	2.274
	<i>Nephroselmis</i> sp. (Al.OTU34)	<i>Crustomastix</i> sp. (Al.OTU100)	0.101	0.156	88	1.387
	<i>Crustomastix</i> sp. (Al.OTU100)	<i>Monomastix</i> sp. (Al.OTU180)	0.156	0.101	88	1.387
Within-group (Fauna)	Unclassified eukaryote (Fa.OTU12)	<i>Mayamaea</i> sp. (Fa.OTU13)	1	1	3	3
	Unclassified eukaryote (Fa.OTU12)	<i>Phascolodon</i> sp. (Fa.OTU23)	1	1	3	3
	<i>Mayamaea</i> sp. (Fa.OTU13)	<i>Phascolodon</i> sp. (Fa.OTU23)	1	1	3	3
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	Unclassified eukaryote (Fa.OTU12)	0	1	0	0
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Mayamaea</i> sp. (Fa.OTU13)	0	1	0	0
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Enchelyodon</i> sp. (Fa.OTU16)	0	1	0	0
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Phascolodon</i> sp. (Fa.OTU23)	0	1	0	0
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Prodorylaimus</i> sp. (Fa.OTU26)	0	0	0	0
	Unclassified eukaryote (Fa.OTU12)	<i>Prodorylaimus</i> sp. (Fa.OTU26)	1	0	0	0
	<i>Mayamaea</i> sp. (Fa.OTU13)	<i>Prodorylaimus</i> sp. (Fa.OTU26)	1	0	0	0

^{a, b} denotes the species keystone after standardizing (Eq. 4)

^c represents the number of togetherness units based on Eq. 9

^d represents their cooperative effects on maximizing the functions calculated by Eq. 12

Table S2C Pairs of core microorganisms and their independent effects on maximizing the temperature sensitivity of N₂O emission in paddy soils based on the two-step criterion.

Interaction type	Node A	Node B	B' (A) ^a	B' (B) ^b	C _{ij} ^c	R _{ij} ^d
Between-group	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Halobacteriovorax</i> sp. (Ba.OTU37210)	0.926	0.831	6,441	4,958.129
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Clostridium</i> sp. (Ba.OTU39051)	0.926	0.834	5,700	4,399.813
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Chondromyces</i> sp. (Ba.OTU6099)	0.926	0.831	5,600	4,308.221
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Oxobacter</i> sp. (Ba.OTU5558)	0.926	0.831	5,424	4,175.017
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Sporotomaculum</i> sp. (Ba.OTU23447)	0.926	0.832	4,180	3,221.426
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Luteitalea</i> sp. (Ba.OTU6324)	0.926	0.830	4,181	3,213.887
	<i>Halobacteriovorax</i> sp. (Ba.OTU37210)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.831	0.842	4,536	3,173.720
	<i>Halobacteriovorax</i> sp. (Ba.OTU37210)	<i>Methermicoccus</i> sp. (Ar.OTU189)	0.831	0.849	4,368	3,083.170
	Unclassified <i>Chondromyces</i> (Ba.OTU6099)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.831	0.842	4,182	2,924.321
	<i>Halobacteriovorax</i> sp. (Ba.OTU37210)	Unclassified <i>Woesearchaeota</i> (Ar.OTU23)	0.831	0.837	4,088	2,842.783
	<i>Clostridium</i> sp. (Ba.OTU39051)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.834	0.842	4,018	2,819.034
Within-group (Archaea)	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Desulfomicrobium</i> sp. (Ba.OTU13113)	0.926	0.832	3,616	2,784.748
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Geobacter</i> sp. (Ba.OTU51222)	0.926	0.830	3,616	2,779.932
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Lysobacter</i> sp. (Ba.OTU100)	0.926	0.836	3,584	2,774.933
	<i>Oxobacter</i> sp. (Ba.OTU5558)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.831	0.842	3,936	2,753.751
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Methanobacterium</i> sp. (Ar.OTU160)	0.818	0.359	713	209.188
	<i>Methanocella</i> sp. (Ar.OTU8)	<i>Methanospaerula</i> sp. (Ar.OTU10)	0.648	0.818	374	198.269
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Methanobacterium</i> sp. (Ar.OTU101)	0.818	0.308	616	155.206

	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Acidianus</i> sp. (Ar.OTU380)	0.818	0.271	612	135.587
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Methanocella</i> sp. (Ar.OTU18)	0.818	0.707	231	133.545
	<i>Methanocella</i> sp. (Ar.OTU22)	<i>Methanobacterium</i> sp. (Ar.OTU93)	1	0.632	210	132.695
Within-group (Bacteria)	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Moorella</i> sp. (Ba.OTU52410)	0.659	1	8,000	5,271.483
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	Unclassified bacterium (Ba.OTU52965)	0.659	0.478	8,256	2,598.729
	<i>Moorella</i> sp. (Ba.OTU52410)	<i>Ammoniphilus</i> sp. (Ba.OTU41498)	1	0.238	6,903	1,645.947
	<i>Moorella</i> sp. (Ba.OTU52410)	<i>Pirellula</i> sp. (Ba.OTU5736)	1	0.477	2,618	1,248.117
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Pyrinomonas</i> sp. (Ba.OTU53333)	0.659	0.442	4,148	1,209.039
	<i>Moorella</i> sp. (Ba.OTU52410)	Unclassified bacterium (Ba.OTU31890)	1	0.198	6,018	1,188.684
	<i>Moorella</i> sp. (Ba.OTU52410)	<i>Thermaanaerothrix</i> sp. (Ba.OTU5016)	1	0.130	8,667	1,128.093
	<i>Moorella</i> sp. (Ba.OTU52410)	<i>Calditerricola</i> sp. (Ba.OTU12669)	1	0.096	11,663	1,115.685
	<i>Moorella</i> sp. (Ba.OTU52410)	Unclassified bacterium (Ba.OTU52965)	1	0.478	2,275	1,086.750
	<i>Frateuria</i> sp. (Ba.OTU45500)	<i>Moorella</i> sp. (Ba.OTU52410)	0.099	1	10,062	995.917
Within-group (Fungi)	Unclassified fungus (Fu.OTU1)	<i>Lobulomyces</i> sp. (Fu.OTU21789)	1	0.301	8	2.407
	Unclassified fungus (Fu.OTU1)	<i>Dominikia</i> sp. (Fu.OTU21962)	1	0.301	8	2.407
	Unclassified fungus (Fu.OTU1)	<i>Cyathus</i> sp. (Fu.OTU10084)	1	0.050	8	0.399
	Unclassified fungus (Fu.OTU1)	Unclassified fungus (Fu.OTU10224)	1	0.067	4	0.270
	<i>Lobulomyces</i> sp. (Fu.OTU21789)	Unclassified Tremellales (Fu.OTU21826)	0.301	0.168	4	0.202
	<i>Dominikia</i> sp. (Fu.OTU21962)	Unclassified Tremellales (Fu.OTU21826)	0.301	0.168	4	0.202
	<i>Rozella</i> sp. (Fu.OTU13113)	Unclassified fungus (Fu.OTU21534)	0.124	0.067	4	0.034
	<i>Rozella</i> sp. (Fu.OTU13113)	Unclassified fungus (Fu.OTU22761)	0.124	0.067	4	0.034
	<i>Rozella</i> sp. (Fu.OTU13113)	<i>Inocybe</i> sp. (Fu.OTU23234)	0.124	0.067	3	0.025
	<i>Rozella</i> sp. (Fu.OTU13113)	<i>Acaulospora</i> sp. (Fu.OTU24067)	0.124	0.067	3	0.025
Within-group (Algae)	<i>Nannochloris</i> sp. (Al.OTU43)	<i>Chloroparvula</i> sp. (Al.OTU52)	0.287	0.907	8	2.080
	<i>Tetracystis</i> sp. (Al.OTU8)	<i>Crustomastix</i> sp. (Al.OTU100)	1	0.116	6	0.695
	<i>Chloroparvula</i> sp. (Al.OTU52)	<i>Pterosperma</i> sp. (Al.OTU388)	0.907	0.251	8	1.823

	<i>Tetracystis</i> sp. (Al.OTU8)	<i>Nephroselmis</i> sp. (Al.OTU34)	1	0.088	6	0.527
	<i>Tetracystis</i> sp. (Al.OTU8)	<i>Monomastix</i> sp. (Al.OTU180)	1	0.088	6	0.527
	<i>Nannochloris</i> sp. (Al.OTU43)	<i>Pterosperma</i> sp. (Al.OTU388)	0.287	0.251	4	0.288
	<i>Nephroselmis</i> sp. (Al.OTU34)	<i>Nautococcus</i> sp. (Al.OTU32)	0.088	0.448	12	0.472
	<i>Monomastix</i> sp. (Al.OTU180)	<i>Nautococcus</i> sp. (Al.OTU32)	0.088	0.448	12	0.472
	<i>Nephroselmis</i> sp. (Al.OTU34)	<i>Monomastix</i> sp. (Al.OTU180)	0.088	0.088	1	0.008
	<i>Nephroselmis</i> sp. (Al.OTU34)	<i>Crustomastix</i> sp. (Al.OTU100)	0.088	0.116	4	0.041
Within-group (Micro-fauna)	Unclassified eukaryote (Fa.OTU12)	<i>Mayamaea</i> sp. (Fa.OTU13)	1	1	1	1
	Unclassified eukaryote (Fa.OTU12)	<i>Phascolodon</i> sp. (Fa.OTU23)	1	1	1	1
	<i>Mayamaea</i> sp. (Fa.OTU13)	<i>Phascolodon</i> sp. (Fa.OTU23)	1	1	1	1
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	Unclassified eukaryote (Fa.OTU12)	0	1	2	0
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Mayamaea</i> sp. (Fa.OTU13)	0	1	2	0
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Mayamaea</i> sp. (Fa.OTU16)	0	1	4	0
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Phascolodon</i> sp. (Fa.OTU23)	0	1	2	0
	<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Prodorylaimus</i> sp. (Fa.OTU26)	0	0	1	0
	Unclassified eukaryote (Fa.OTU12)	<i>Prodorylaimus</i> sp. (Fa.OTU26)	1	0	2	0
	<i>Mayamaea</i> sp. (Fa.OTU13)	<i>Prodorylaimus</i> sp. (Fa.OTU26)	1	0	2	0

^{a, b} denotes the species keystone ness after standardizing (Eq. 4)

^c represents the number of checkboard units based on Eq. 7

^d represents their independent effects on maximizing the functions calculated by Eq. 11

Table S2D Pairs of core microorganisms and their cooperative effects on maximizing the temperature sensitivity of N₂O emission in paddy soils based on the two-step criterion.

Interaction type	Node A	Node B	B' (A) ^a	B' (B) ^b	T _{ij} ^c	R _{ijt} ^d
Between-group	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Methanobacterium</i> sp. (Ar.OTU13)	0.958	1	54,696	52,390.998
	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Methanocella</i> sp. (Ar.OTU32)	0.958	0.920	53,193	46,877.402
	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Nitrososphaera</i> sp. (Ar.OTU9)	0.958	0.924	46,530	41,185.715
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Nautococcus</i> sp. (Al.OTU32)	0.926	0.865	50,550	40,507.698
	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Methanocella</i> sp. (Ar.OTU22)	0.958	0.962	40,677	37,487.655
	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Nitrososphaera</i> sp. (Ar.OTU4)	0.958	0.895	42,517	36,443.019
	<i>Nautococcus</i> sp. (Al.OTU32)	Unclassified Archaeon (Ar.OTU135)	0.865	0.847	47,790	35,034.359
	<i>Nautococcus</i> sp. (Al.OTU32)	<i>Methanocella</i> sp. (Ar.OTU143)	0.865	0.865	44,058	32,984.802
	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Methanocella</i> sp. (Ar.OTU18)	0.958	0.892	33,216	28,385.398
	<i>Nautococcus</i> sp. (Al.OTU32)	<i>Methanospaerula</i> sp. (Ar.OTU36)	0.865	0.856	37,908	28,081.625
Within-group (Archaea)	<i>Nautococcus</i> sp. (Al.OTU32)	<i>Methanococcoides</i> sp. (Ar.OTU74)	0.865	0.838	29,708	21,536.073
	<i>Nautococcus</i> sp. (Al.OTU32)	Unclassified Archaeon (Ar.OTU531)	0.865	0.832	25,656	18,462.526
	<i>Chlamydomonadales</i> sp. (Al.OTU16)	<i>Methanosarcina</i> sp. (Ar.OTU98)	0.958	0.856	22,218	18,227.753
	<i>Frontonia</i> sp. (Fu.OTU23455)	Unclassified Archaeon (Ar.OTU23)	0.852	0.837	25,416	18,108.302
	<i>Frontonia</i> sp. (Fu.OTU23455)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.852	0.842	25,224	18,082.032
	<i>Methermicoccus</i> sp. (Ar.OTU189)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.478	0.491	9,065	2,129.871
	<i>Methanospaerula</i> sp. (Ar.OTU10)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.818	0.491	5,220	2,096.382

	<i>Methanobacterium</i> sp. (Ar.OTU125)	<i>Methanobacterium</i> sp. (Ar.OTU1004)	0.336	0.491	8,460	1,394.688
	<i>Methanosphaerula</i> sp. (Ar.OTU10)	Unclassified <i>Woesearchaeota</i> (Ar.OTU23)	0.818	0.366	4,500	1,348.396
	Unclassified <i>Woesearchaeota</i> (Ar.OTU23)	<i>Methermicoccus</i> sp. (Ar.OTU189)	0.366	0.478	7,644	1,340.020
Within-group (Bacteria)	<i>Moorella</i> sp. (Ba.OTU52410)	Unclassified <i>bacterium</i> (Ba.OTU52965)	1	0.478	79,106	37,788.342
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Moorella</i> sp. (Ba.OTU52410)	0.659	1	39,936	26,315.246
	<i>Moorella</i> sp. (Ba.OTU52410)	<i>Gemmata</i> sp. (Ba.OTU8890)	1	0.251	83,006	20,862.333
	<i>Moorella</i> sp. (Ba.OTU52410)	<i>Calditerricola</i> sp. (Ba.OTU1225)	1	0.236	74,550	17,613.370
	<i>Moorella</i> sp. (Ba.OTU52410)	<i>Sterolibacterium</i> sp. (Ba.OTU11112)	1	0.192	85,162	16,349.244
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Pyrinomonas</i> sp. (Ba.OTU53333)	0.659	0.442	54,213	15,801.748
	<i>Moorella</i> sp. (Ba.OTU52410)	<i>Chryseobacterium</i> sp. (Ba.OTU14780)	1	0.252	44,321	11,189.741
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Denitratisoma</i> sp. (Ba.OTU51233)	0.659	0.240	70,528	11,171.385
	<i>Anaerosalibacter</i> sp. (Ba.OTU41112)	<i>Peredibacter</i> sp. (Ba.OTU54965)	0.659	0.193	81,770	10,393.831
	<i>Moorella</i> sp. (Ba.OTU52410)	<i>Pirellula</i> sp. (Ba.OTU5736)	1	0.477	21,640	10,316.748
Within-group (Fungi)	<i>Rozella</i> sp. (Fu.OTU13113)	<i>Inocybe</i> sp. (Fu.OTU23234)	0.124	0.067	26	0.218
	<i>Rozella</i> sp. (Fu.OTU13113)	<i>Acaulospora</i> sp. (Fu.OTU24067)	0.124	0.067	26	0.218
	<i>Coemansia</i> sp. (Fu.OTU7779)	Unclassified <i>fungus</i> (Fu.OTU13335)	0.067	0.067	28	0.127
	<i>Coemansia</i> sp. (Fu.OTU7779)	Unclassified <i>Cryptomycota</i> (Fu.OTU21900)	0.067	0.067	28	0.127
	Unclassified <i>fungus</i> (Fu.OTU13335)	Unclassified <i>Cryptomycota</i> (Fu.OTU21900)	0.067	0.067	28	0.127
	<i>Inocybe</i> sp. (Fu.OTU23234)	Unclassified <i>fungus</i> (Fu.OTU24067)	0.067	0.067	28	0.127
	Unclassified <i>Metazoa</i> (Fu.OTU1)	<i>Lobulomyces</i> sp. (Fu.OTU21789)	1	0.301	0	0
	Unclassified <i>Metazoa</i> (Fu.OTU1)	<i>Dominikia</i> sp. (Fu.OTU21962)	1	0.301	0	0
	Unclassified <i>Metazoa</i> (Fu.OTU1)	<i>Cyathus</i> sp. (Fu.OTU10084)	1	0.050	0	0
	Unclassified <i>Metazoa</i> (Fu.OTU1)	Unclassified <i>fungus</i> (Fu.OTU10224)	1	0.067	0	0
Within-group (Algae)	<i>Nannochloris</i> sp. (Al.OTU43)	<i>Chloroparvula</i> sp. (Al.OTU52)	0.287	0.907	43	11.178
	<i>Tetraclysis</i> sp. (Al.OTU8)	<i>Crustomastix</i> sp. (Al.OTU100)	1	0.116	86	9.957
	<i>Chloroparvula</i> sp. (Al.OTU52)	<i>Pterosperma</i> sp. (Al.OTU388)	0.907	0.251	43	9.796

<i>Tetracystis</i> sp. (Al.OTU8)	<i>Nephroselmis</i> sp. (Al.OTU34)	1	0.088	86	7.550
<i>Tetracystis</i> sp. (Al.OTU8)	<i>Monomastix</i> sp. (Al.OTU180)	1	0.088	86	7.550
<i>Nannochloris</i> sp. (Al.OTU43)	<i>Pterosperma</i> sp. (Al.OTU388)	0.287	0.251	45	3.240
<i>Nephroselmis</i> sp. (Al.OTU34)	<i>Nautococcus</i> sp. (Al.OTU32)	0.088	0.448	42	1.650
<i>Monomastix</i> sp. (Al.OTU180)	<i>Nautococcus</i> sp. (Al.OTU32)	0.088	0.448	42	1.650
<i>Nephroselmis</i> sp. (Al.OTU34)	<i>Monomastix</i> sp. (Al.OTU180)	0.088	0.088	135	1.041
<i>Nephroselmis</i> sp. Al.OTU34	<i>Crustomastix</i> sp. Al.OTU100	0.088	0.116	88	0.895
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Within-group (Micro-fauna)					
Unclassified eukaryote (Fa.OTU12)	<i>Mayamaea</i> sp. (Fa.OTU13)	1	1	3	3
Unclassified eukaryote (Fa.OTU12)	<i>Phascolodon</i> sp. (Fa.OTU23)	1	1	3	3
<i>Mayamaea</i> sp. (Fa.OTU13)	<i>Phascolodon</i> sp. (Fa.OTU23)	1	1	3	3
<i>Aplanochytrium</i> sp. (Fa.OTU7)	Unclassified eukaryote (Fa.OTU12)	0	1	0	0
<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Mayamaea</i> sp. (Fa.OTU13)	0	1	0	0
<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Enchelyodon</i> sp. (Fa.OTU16)	0	1	0	0
<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Phascolodon</i> sp. (Fa.OTU23)	0	1	0	0
<i>Aplanochytrium</i> sp. (Fa.OTU7)	<i>Prodorylaimus</i> sp. (Fa.OTU26)	0	0	0	0
Unclassified eukaryote (Fa.OTU12)	<i>Prodorylaimus</i> sp. (Fa.OTU26)	1	0	0	0
<i>Mayamaea</i> sp. (Fa.OTU13)	<i>Prodorylaimus</i> sp. (Fa.OTU26)	1	0	0	0

^{a, b} denotes the species keystone ness after standardizing (Eq. 4)

^c represents the number of togetherness units based on Eq. 9

^d represents their cooperative effects on maximizing the functions calculated by Eq. 12